



0960-894X(94)00222-3

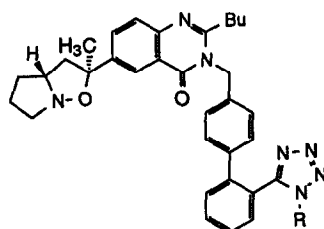
**Synthesis and Biological Evaluation of the Potent
Isoxazolidinyl Angiotensin II Receptor Antagonist
CL332,877 and Its Enantiomers.**

Jeremy I. Levin,* Peter S. Chan, Joseph Coupet, Lucien Thibault
A. M. Venkatesan, Trina K. Bailey, George Vice,
Agnes Cobuzzi, Fong Lai, Noel Mellish

*American Cyanamid Company, Medical Research Division,
Lederle Laboratories, Pearl River,
NY 10965*

Abstract: An alternative synthesis of CL332,877, a potent isoxazolidinyl-quinazolinone angiotensin II receptor antagonist, is described. In addition, the enantiomers of CL332,877 were separated and evaluated both *in vitro* and *in vivo*.

In the course of our work on quinazolinone based angiotensin II (A II) receptor antagonists we prepared a series of 6-isoxazolidinyl derivatives, from which CL332,877 (**1a**) was chosen for pre-clinical development as an agent for the treatment of hypertension.¹ CL332,877 was found to be an exceptionally potent, long-acting, orally active, non-competitive A II antagonist. However, two major

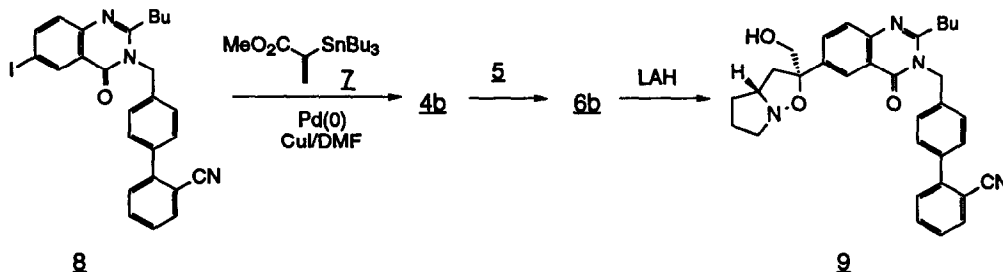


1a R = Na CL332,877

1b R = H CL190,133

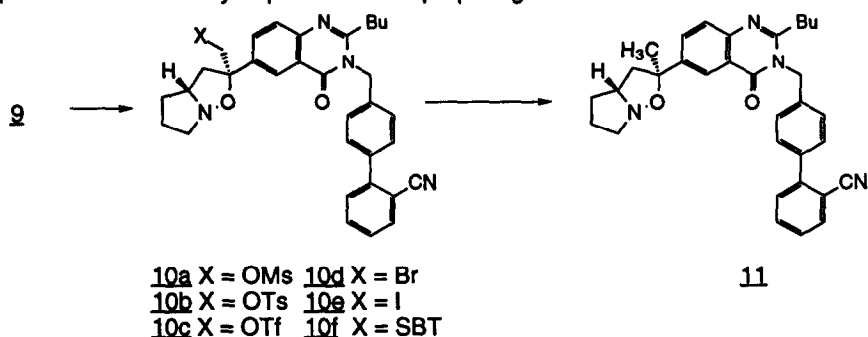
problems presented barriers to the timely development of this compound. First, the isoxazolidine moiety of CL332,877 is derived from the minor product of the diastereomeric pair produced in a nitron cycloaddition with alkenyl-quinazolinone **2** (Scheme 1). The desired diastereomer **3a** must be separated by a difficult, tedious chromatography from the undesired major isomer **3b** and is obtained in only 20-25% yield under optimum conditions. Second, since CL332,877 is synthesized in racemic form, a route was needed to gain access to each enantiomer in order to evaluate their individual biological properties. We now report solutions to both of these problems along with the results of *in vitro* and *in vivo* screens assessing the activity of each of the enantiomers of CL332,877 as A II receptor antagonists.

functionality into the methyl group of CL332,877 without cleaving the sensitive N-O bond of the isoxazolidine ring.



Scheme 3

Several methods were investigated for removal of the hydroxyl functionality from **9** (Scheme 4). Initially, alcohol **9** was converted into the corresponding neopentyl mesylate **10a** or tosylate **10b**, followed by attempted reduction to the alkane with various hydride reagents (e.g. LAH, LiHBEt₃, etc.).⁴ These procedures returned starting material under normal conditions and, under more forcing conditions, gave products resulting from reduction of the quinazolinone or isoxazolidine rings. Even the triflate derivative **10c** could not be converted into the alkane under an assortment of conditions (e.g. H₂, LAH/TiCl₃, NaBH₃CN/DMF, NaBH₄/t-BuOH, Li₅CuH₆, Na/NH₃, Ra Ni).⁴ Next, alcohol **9** was converted into bromide **10d** in 60% yield, using 1,2-dibromotetrachloroethane/Bu₃P,⁵ and then subjected to a wide variety of reducing agents without success. Radical initiated reduction of **10d** with tri-n-butyltin hydride did give a product resulting from reduction of the bromide, but the bicyclic isoxazolidine had apparently rearranged in the process, possibly by insertion of the methyl radical into the isoxazolidine ring. The same problem was encountered on attempted radical deoxygenation of thiocarbonyl derivatives of **9** using numerous procedures described by Barton.⁶ Iodide **10e**, synthesized in 30% yield from alcohol **9** (I₂, Bu₃P, Bu₄N⁺I⁻), proved to be similarly unproductive for preparing the desired alkane.

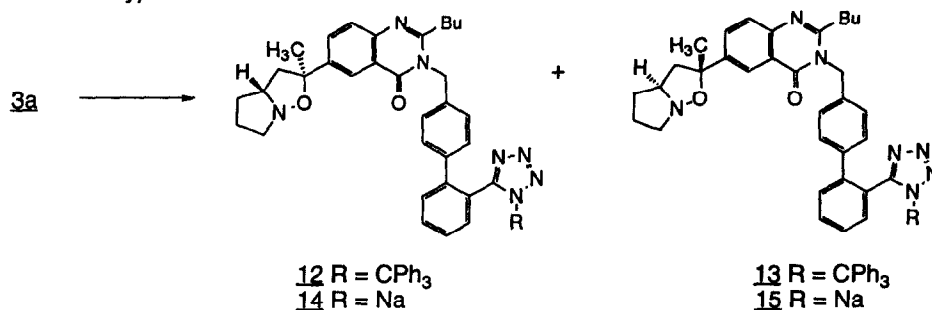


Scheme 4

Finally it was determined that alcohol **9** could be converted into sulfide **10f** (SBT = 2-S-benzothiazole) efficiently by reaction with 2,2'-dithiobis(benzothiazole) and tributylphosphine in

refluxing toluene.⁷ Careful room temperature desulfurization of **10f** using activated Raney nickel in DMF, with a small amount of added pyridine, gave the desired angular methyl compound **11**, with minimal cleavage of the isoxazolidine ring (Scheme 4).⁸ Numerous other desulfurization techniques were attempted with no improvement over this procedure. The overall yield for conversion of isoxazolidine-ester **8** to compound **11** was 30%, and no chromatography was required. Biphenyl nitrile **11** was then converted into trityl tetrazole **3a** by the procedure of Duncia⁹, followed by deprotection and formation of the tetrazole sodium salt to give **1a** in 74% yield.

Although the synthetic sequence described above could conceivably be adapted to provide an asymmetric synthesis of either enantiomer of CL332,877 by using a chiral acrylamide derivative rather than acrylate ester **4b**, a more expedient method was pursued. Thus, chromatography of trityl tetrazole derivative **3a** on a chiral column (Chiracel OD-R) eluting with hexanes/isopropanol (7:3) gave each of the enantiomers, **12** and **13**, in greater than 99% e.e. (Scheme 5). This technique was successfully used to prepare ~100mg of each of the two enantiomers which were then detritylated and converted into their respective sodium salts, **14** and **15** (absolute configurations have been assigned arbitrarily).



Scheme 5

Biological evaluation of the enantiomers was performed both *in vitro* and *in vivo*. Binding affinities of the enantiomers were measured using both bovine adrenal cortex and rat adrenal cortex.¹⁰ IC₅₀ values for racemate **1a**, the enantiomers **14** and **15**, and DuP753 (losartan) are shown in Table 1. Thus, the racemic parent compound, **1a**, and enantiomer **14** are approximately two-fold more potent than enantiomer **15**, and at least three-fold more potent than DuP753.

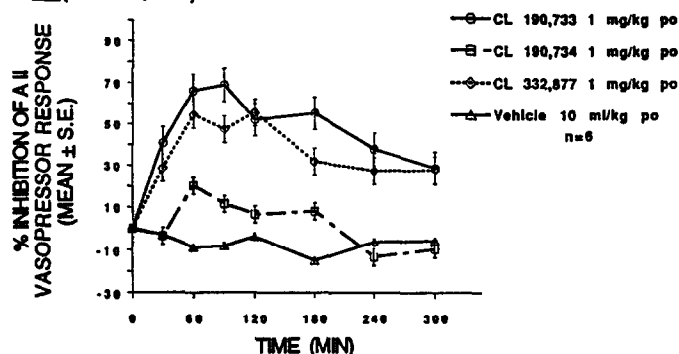
Table 1: Relative IC₅₀ Values for **1a**, **14**, **15** and DuP753.

Tissue Type	DuP 753	1a	14	15
Rat adrenal	57 nM	17 nM	17 nM	26 nM
Bovine "	330 nM	26 nM	38 nM	77nM

The same two-fold difference in potency between the enantiomers is seen *in vivo* in an A II challenge screen measuring the antagonism of the vasopressor response to a 0.05 µg/kg i.v. dose

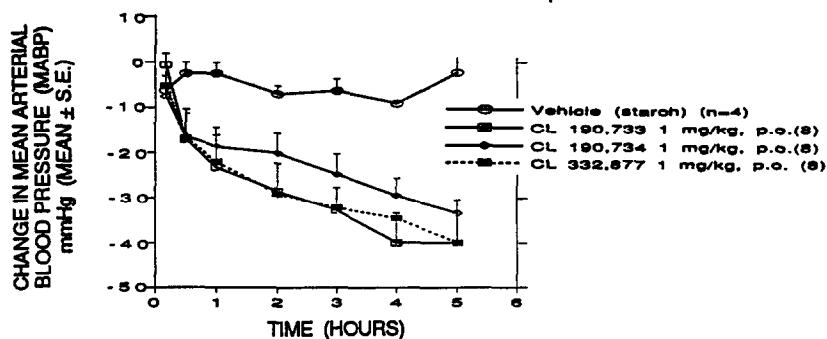
of A II in conscious spontaneously hypertensive rats (SHR) (Figure 1).¹¹ Enantiomer **14** is slightly more potent than racemate **1a** in this assay.

Figure 1: Inhibition of A II Challenge in SHR by a 1mg/kg Oral Dose of **1a**(CL332,877), **14**(CL190,733) and **15**(CL190,734).



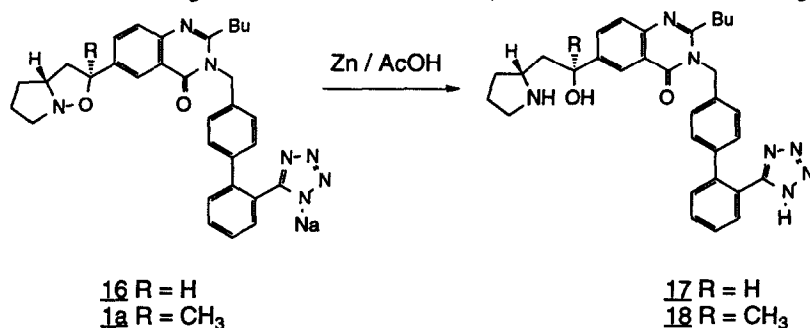
On the other hand, on oral dosing in an aorta-coarcted rat model¹¹ of renin dependent hypertension compounds **14** and racemate **1a** were equipotent, both being approximately twice as active as enantiomer **15**, (Figure 2). The enantiomers were further characterized by their effect on the A II-induced contraction of rabbit femoral artery. Schild plot analysis of this data determined that both **14** and **15** were non-competitive antagonists of A II with apparent pA_2 's of 10.6 and 10.7 respectively (**1a** has previously been shown¹ to be a non-competitive antagonist with a $pA_2 = 10.9$).

Figure 2: Effect of Oral Administration of **1a**(CL332,877), **14**(CL190,733) and **15**(CL190,734) (1mg/kg) on Mean Arterial Blood Pressure in Conscious Renin-Dependent Aorta-Coarcted Rats.



It should be noted at this point that **1a** was also tested for its metabolic stability, since we had previously determined that compound **16**¹ was rapidly and extensively metabolized on incubation with hepatic subcellular fractions from both rat and man.^{12a} One of the metabolites was identified as the ring-opened amino-alcohol **17** (Scheme 6).^{12b} The putative metabolite of **1a** was synthesized by reaction with zinc in acetic acid to provide **18** in 41% yield. Amino-alcohol **18** proved to be an

extremely potent A II antagonist *in vitro* (IC_{50} = 3.5nM), ten times more potent than Merck's imidazo-[4,5-b]pyridine A II antagonist L-158,809¹³ (IC_{50} = 27nM) in the same assay.¹⁰ However, **18** was only moderately active on oral dosing (3mg/kg p.o.) in the aorta-coarcted rat model of hypertension, causing a maximum 32mm Hg fall in mean arterial blood pressure 4 hours after dosing.



Scheme 6

In summary, we have developed effective synthetic routes to the potent A II receptor antagonist CL332,877 (**1a**) each of its enantiomers (**14** and **15**), and a possible metabolite (**18**). Compounds **1a**, **14**, **15** and **18** were all found to be extremely potent *in vitro* while significant differences were found on oral dosing in two *in vivo* models for A II receptor antagonist activity.

References and Notes:

- Levin, J. I.; Chan, P. S.; Coupet, J.; Bailey, T. K.; Vice, G.; Thibault, L.; Lai, F.; Venkatesan, A. *M. Bioorg. & Med. Chem. Lett.* submitted.
- For a review of nitro cycloadditions see: Tufariello, J. J. *1,3-Dipolar Cycloaddition Chemistry*, Padwa, A. Ed.; J. Wiley & Sons, 1984; Vol. 2; pp.83-168.
- Levin, J. I. *Tet. Lett.* **1993**, *34*, 6211.
- For a list of methods for reductions to provide alkanes see: Larock, R. C. *Comprehensive Organic Transformations*, VCH Publishers, 1989; pp.18-20 & 27-35.
- Bringmann, G.; Schneider, S. *Synthesis* **1983**, 139.
- Barton, D. H. R.; Jang, D. O.; Jaszberenyi, J. C. *Tet. Lett.* **1991**, *33*, 5709 and references therein.
- Watanabe, Y.; Araki, T.; Ueno, Y.; Endo, T. *Tet. Lett.* **1986**, *27*, 5383.
- Petit, G. R.; van Tamelen, E. E. *Organic Reactions* **1962**, *12*, 356.
- Duncia, J. V.; Pierce, M. E.; Santella, J. B. *J. Org. Chem.* **1991**, *56*, 2395.
- a) Chiu, A. T.; Duncia, J. V.; McCall, D. E.; Wong, P. C.; Price, W. A.; Thoolen, M. J. N. C.; Carini, D. J.; Johnson, A. L.; Timmermans, P. B. M. W. M. *J. Pharmacol. Exper. Therap.* **1989**, *250*, 867.
b) Assay done using bovine adrenal cortex.
- Chan, P. S.; Cervoni, P.; Ronsberg, M. A.; Scully, P. A.; Quirk, G. J.; McLendon, T. K. *Drug Development Res.* **1989**, *18*, 75.
- a) Dr. M. Kohlbrenner, unpublished results.
b) Dr. W. DeMaio, unpublished results.
- Mantlo, N. B.; Chakravarty, P. K.; Ondeyka, D. L.; Siegl, P. K. S.; Chang, R. S.; Lotti, V. J.; Faust, K. A.; Chen, T.-B.; Schorn, T. W.; Sweet, C. S.; Emmert, S. E.; Patchett, A. A.; Greenlee, W. J. *J. Med. Chem.* **1991**, *34*, 2922.

(Received in USA 9 May 1994; accepted 7 June 1994)